

Transcranial electrical stimulation modulates firing rates at clinically relevant intensities

Farahani et al (2024)

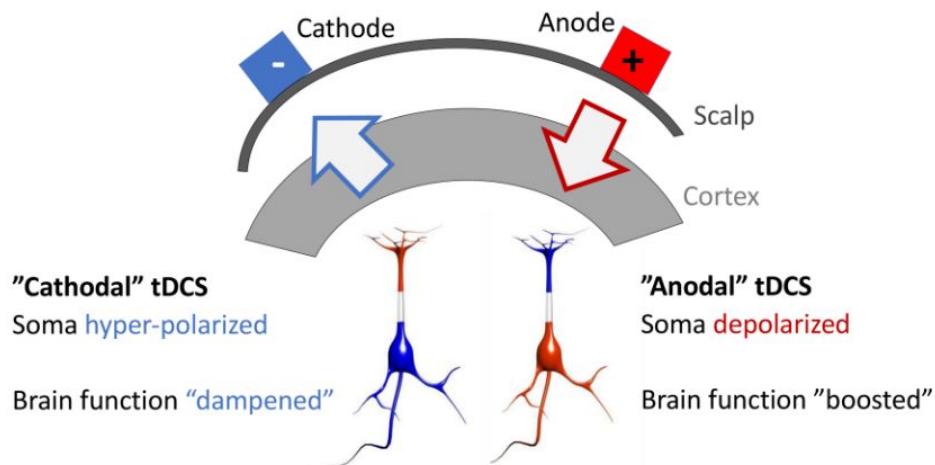
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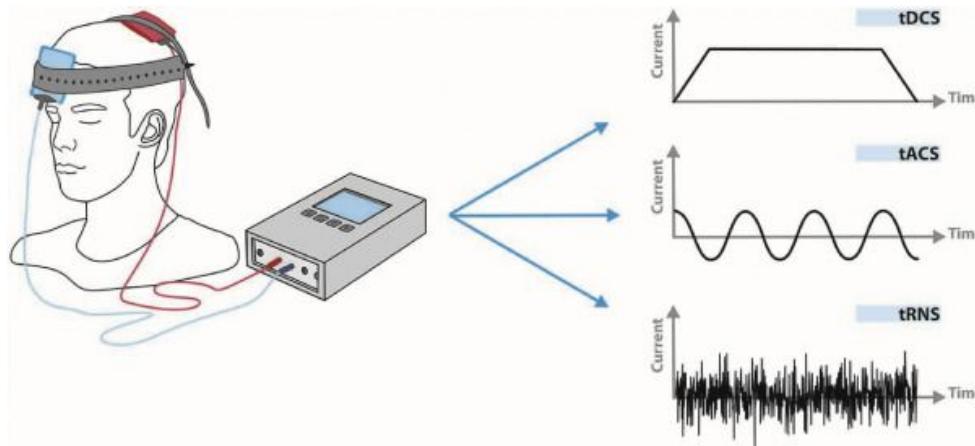
Introduction

Background on transcranial
electrical stimulation and
motivation for the study

- Non-invasive neuromodulation technique
- 2 electrodes on the skin generating electric fields, affecting neuronal membranes by acting on their polarization
- Excite or inhibit depending on the current

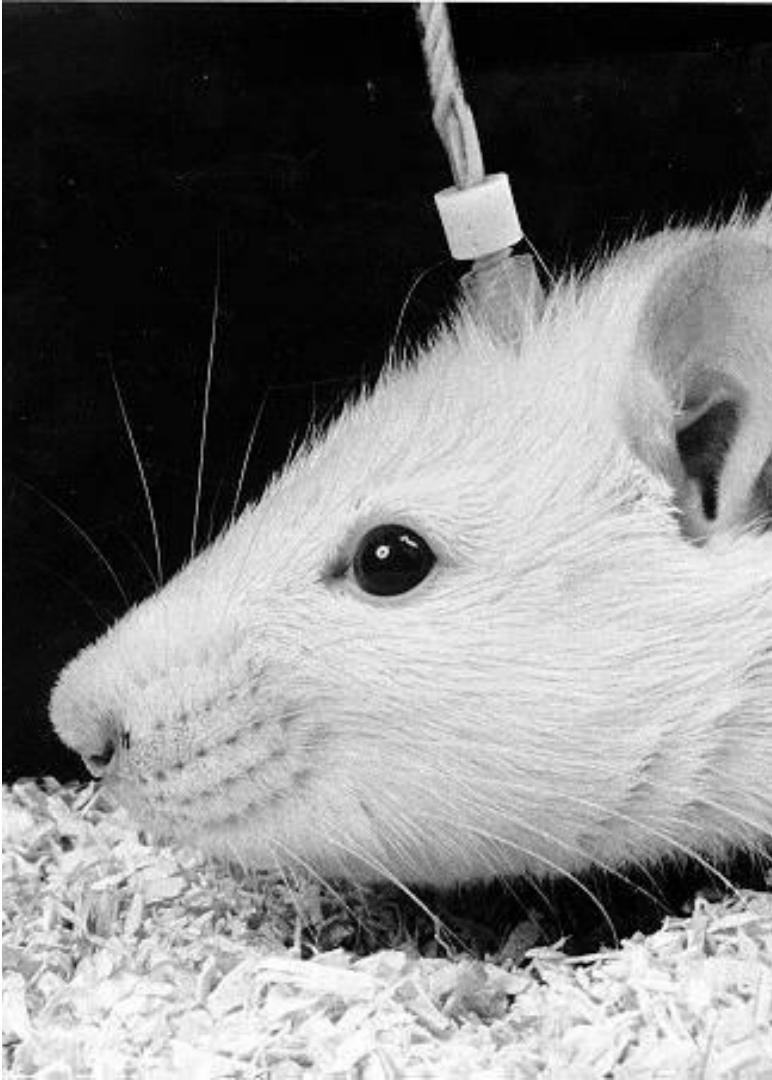


- Transcranial direct current (tDCS), alternating current (tACS) or random noise (tRNS) stimulation



- Problem: remains a critical gap in understanding efficacy and impact of low-intensity electric fields within a living brain
 - significant gap between electric fields
 - **in vivo in human** brain ($\leq 0.5 \text{ V/m}$)
 - **in vitro animal** experiments ($\geq 5 \text{ V/m}$)
- Question: can the low field intensities used in clinical settings induce measurable, relevant changes in neuronal behavior ?

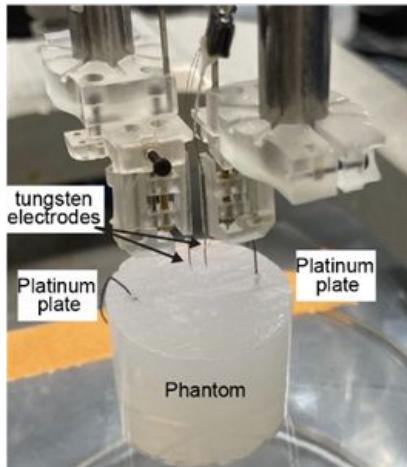
- Goal of study: investigate the effects of clinically relevant electric field strengths on hippocampal neurons in freely moving rats
 1. Measure **electric fields magnitude** and their effects on neuronal firing rate *in vivo* in rats
 2. Establish **calibrated computational models of current flow** in motor cortex and hippocampus



Methods

Techniques and methods used to ensure robustness of the study

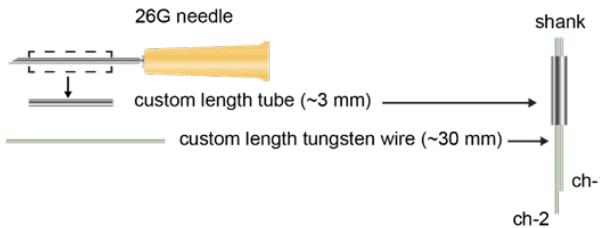
Experiment preparation



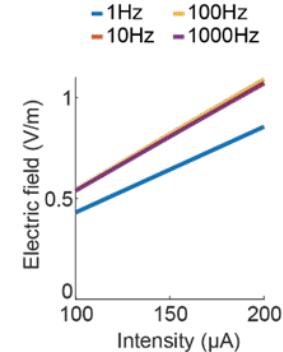
Equipment calibration with phantom brain

- Agar solution with T1 and T2 relaxation similar to gray matter
- Different frequencies (1, 10, 100, 1000 Hz)
- Different intensities (100, 150, 200 μ A)
- Linear increase of electric field magnitude with stimulation intensity

Custom tungsten recording electrode



- 4 channel, 2 shank device
- Record electric field magnitude
- Attached to mechanical shuttle (microdrive)



Computational models

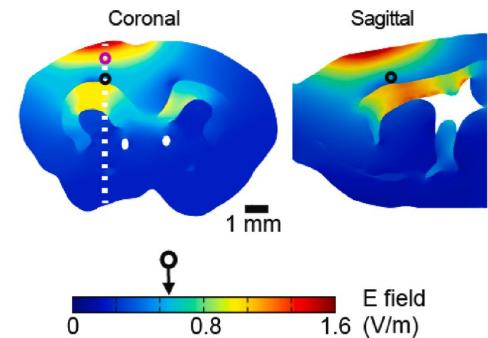
Electric field prediction

- Conductor model based on segmented MRI rat head scan
- Electrodes placed according to experiment coordinates
- Mesh of rat model imported in COMSOL

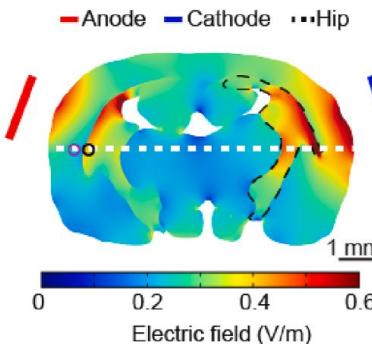


- Finite element analysis software
- Literature based conduction values for each head compartment
- 2D simulation of brain slice

Montage 1:

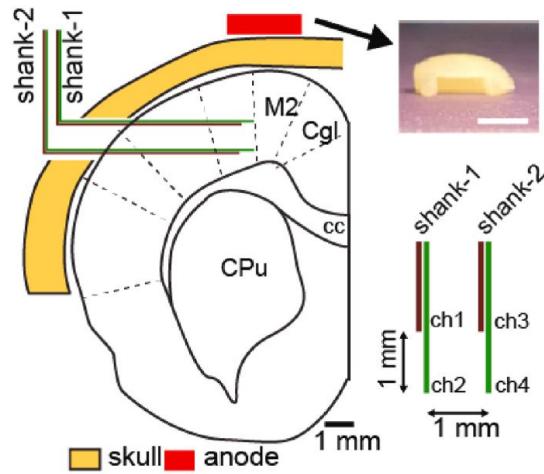


Montage 2:

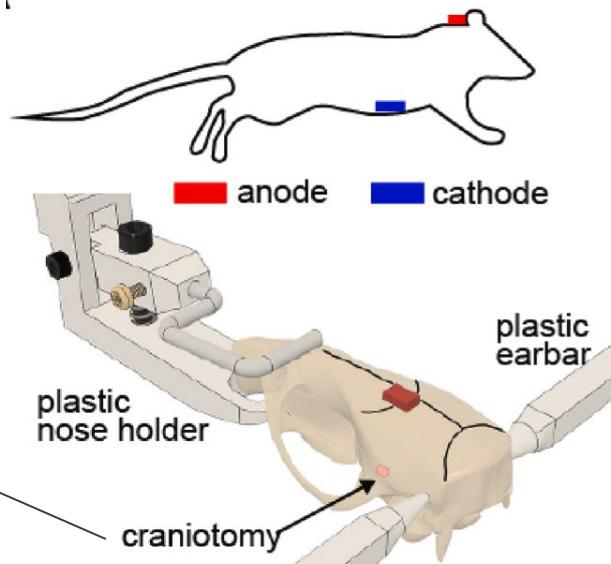


Experimental setup on anesthetized rats (montage 1)

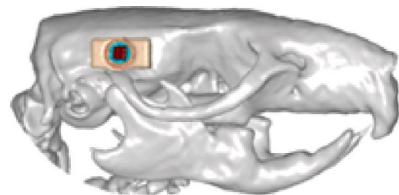
- Motor cortex stimulation and recording
- Varying frequency (10, 100, 1000 Hz)
- Varying intensity (10, 20, 40 μ A)



Montage 1 electrode placement and setup:



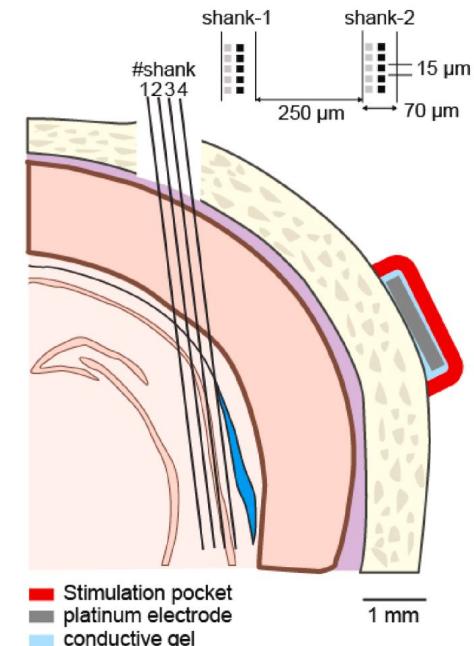
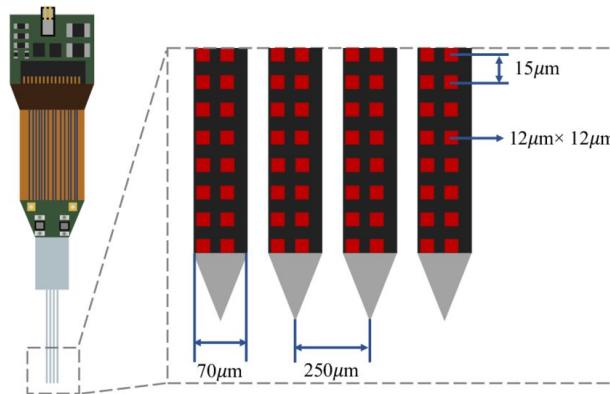
Experimental setup on freely moving rats (montage 2)



- Symmetrical (sagittal plane) electrode placement on temporal bone
- Varying frequency (10, 100, 1000Hz)
- Varying intensity (10, 20, 30 μ A)
- Direct current for single unit recording (3-4s bursts)

Neuropixels 2.0:

- 4 shanks
- 5120 recording sites
- 384 channels per probe
- Implanted in intermediate CA2 region

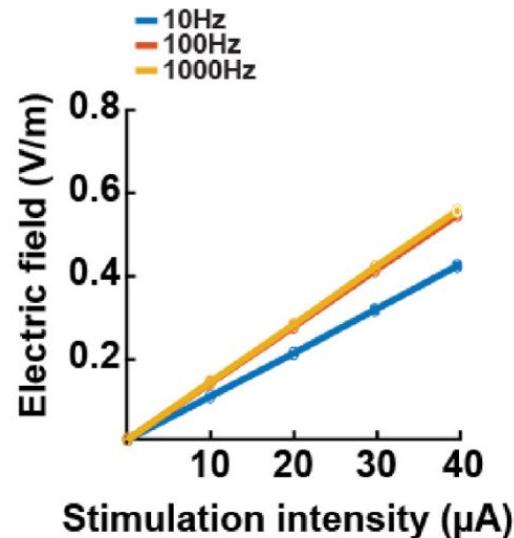




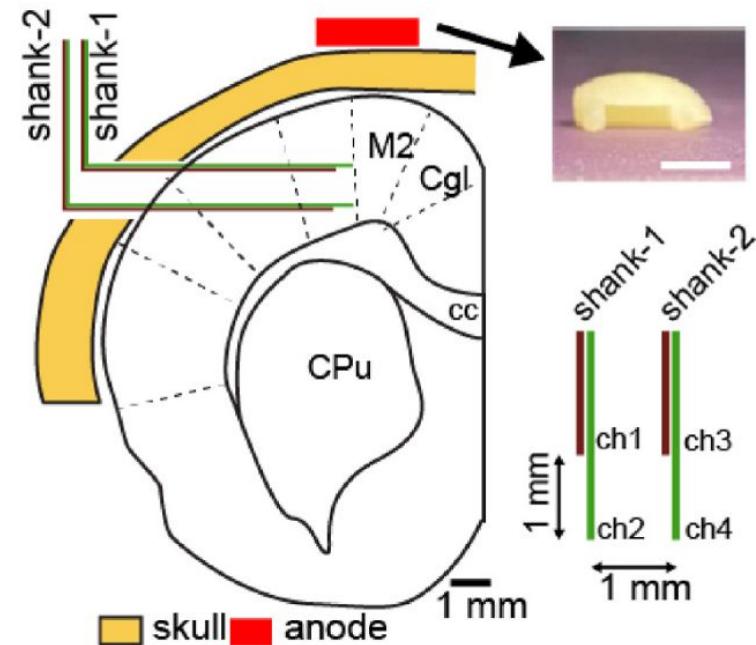
Results

Presentation and analysis of
experiment results

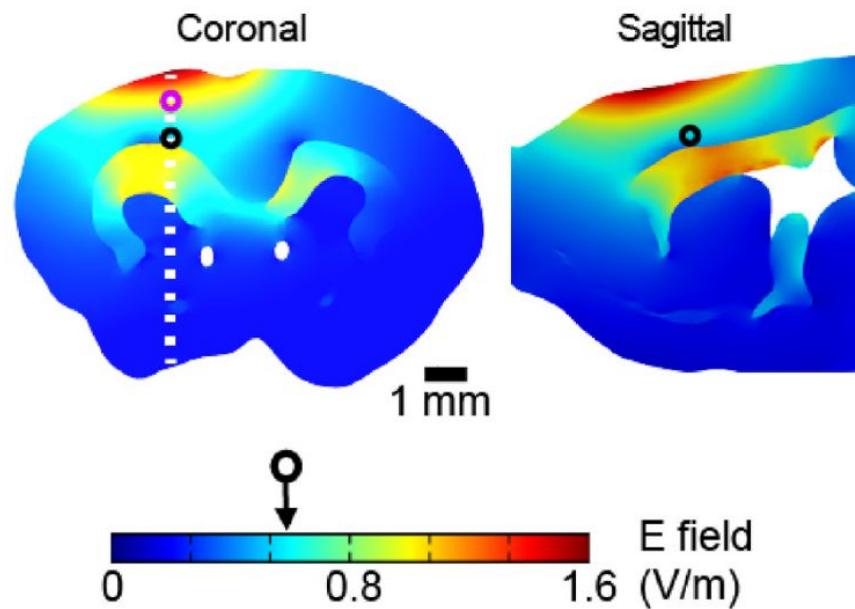
Electric field magnitude increases linearly with applied current in the motor cortex



Slope average on all three = 15 V/m



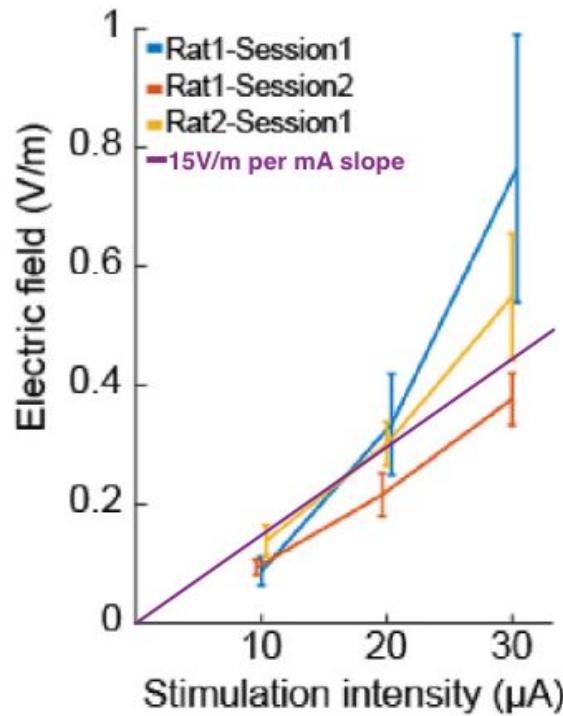
Computational model results are consistent with in-vivo results in the motor cortex



Model prediction in motor cortex at $40 \mu\text{A}$ current:
 0.602 V/m

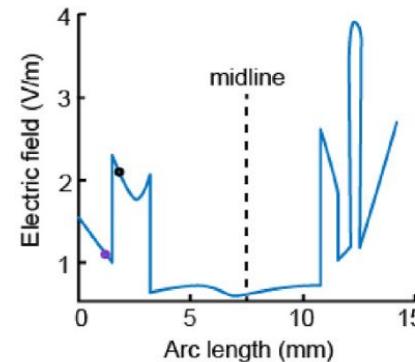
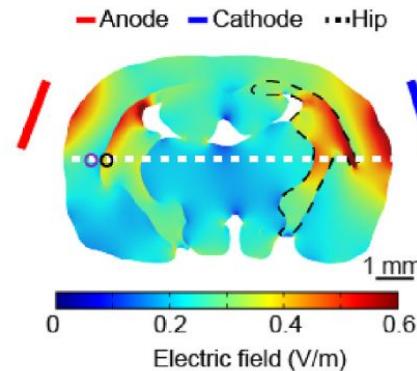
This is equal to 15.05 V/m per mA

Electric field magnitude increases approximately linearly with applied current in the hippocampus



- The slope corresponds to 10, 14 and 18.7 V/m per mA (average = 14.23 V/m per mA)
- Variation in tissue conductivity and structure between hippocampus and cortex leads to weaker electric field for the same amount of current.
- They assume 15 V/m per mA in the hippocampus for the remainder of the paper

Computational model results are approximately consistent with in-vivo results in the hippocampus

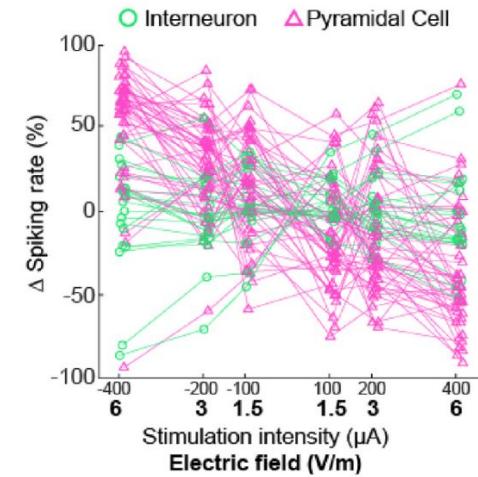
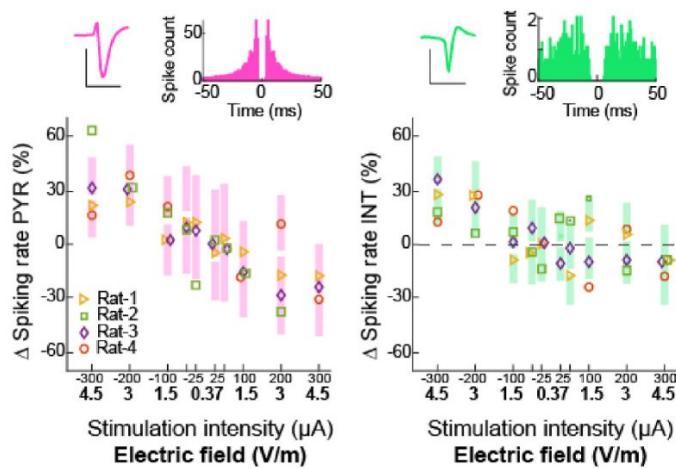
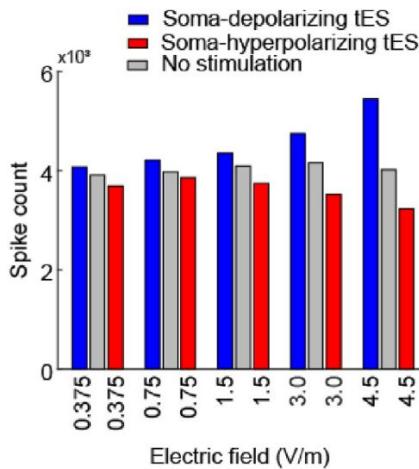


Model prediction in white matter: 2.1 V/m (= 21V/m per mA)

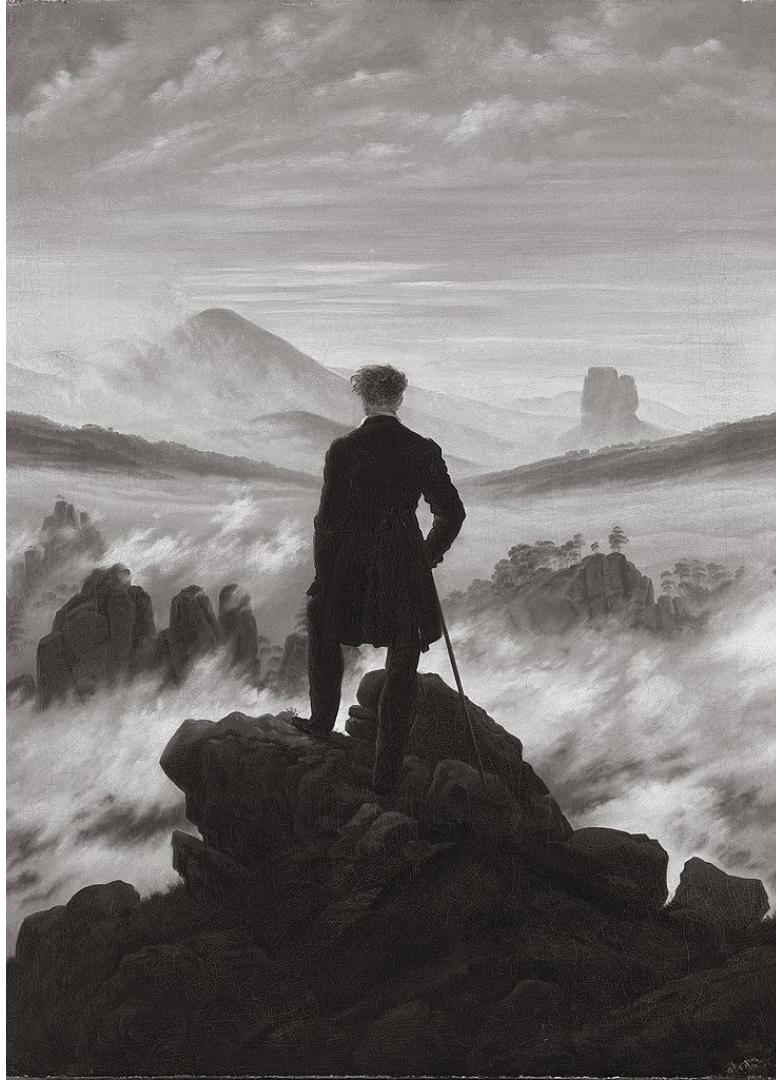
Model prediction in grey matter: 1.2 V/m (= 12 V/m per mA)

Anatomical structure influenced the distribution of electric fields across the hippocampus (discontinuity in conductance between white and grey matter)

Polarity-dependent changes in firing rates in the hippocampus



- The firing rate of excitatory neurons increased with soma-depolarizing stimulation and decreased with soma-hyperpolarizing stimulation

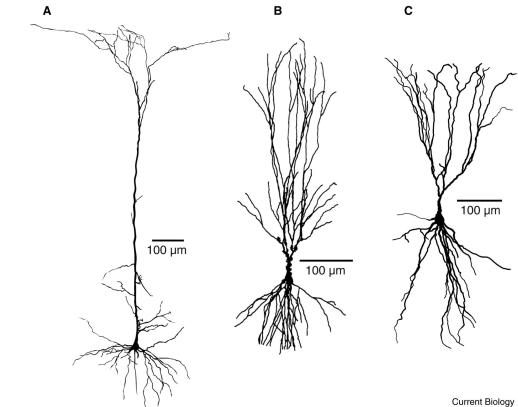


Discussion

Contribution of the study to our understanding of transcranial electrical stimulation

Key findings:

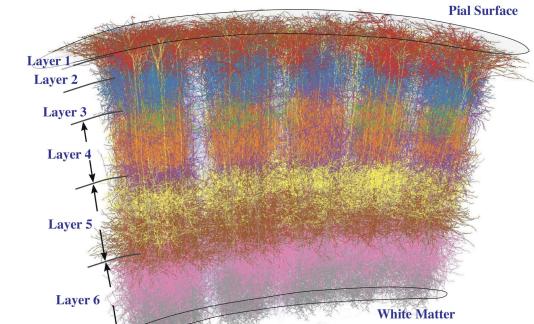
- Low intensity electric fields (~0.5 V/m) can modulate neuronal firing rates *in vivo* at clinically relevant levels
- Pyramidal neuron orientation make them particularly sensitive to field orientation
- Interneurons have a more varied response, likely due to network connectivity



Current Biology

Implications for clinical tES:

- Aligning field orientation with cell structures in specific brain regions optimizes neuromodulatory impact
- Researching further synaptic plasticity response at low-intensity fields could guide tES paradigms for clinical application



3D reconstruction of five columns in rat vibrissal cortex

underlying image from:
Marcel Oberländer, Beyond the Cortical Column, Neuroinformatics 2012

Bridging the gap between animal and human study



Estimates suggest field intensities can be ten times stronger in animals compared to humans



The study suggests for any animal tES study:

- Measuring electric field using sinusoidal waveforms at three different intensities
- Calibrating equipment using a phantom

Short term focus on acute effects:

Limitation: The study measured neuronal firing rates during short bursts of electrical stimulation (3-4s)

Challenge: Explore cumulative effect of repeated tES stimulation on synaptic plasticity

Focus on single brain region (Hippocampus):

Limitation: Neuronal orientation varies across brain regions, effects observed may not translate to other regions

Challenge: Extending experiments to other brain regions to generalize findings

Other possible directions:

- Integrating behavioral tests
- Increase sample size
- Human translation



Transcranial electric stimulation modulates firing rate at clinically relevant intensities

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ABSTRACT

Background: Notwithstanding advances with low-intensity transcranial electrical stimulation (tES), there remain questions about the efficacy of clinically realistic electric fields on neuronal function.

Objective: To measure electric fields magnitude and their effects on neuronal firing rate of hippocampal neurons in freely moving rats, and to establish calibrated computational models of current flow.

Methods: Current flow models were calibrated on electric field measures in the motor cortex ($n = 2$ anesthetized rats) and hippocampus. A Neuroptekels 2.0 probe with 384 channels was used in an *in-vivo* rat model of tES ($n = 4$ freely moving and 2 urethane anesthetized rats) to detect effects of weak fields on neuronal firing rate. High-density field mapping and computational models verified field intensity (1 V/m in hippocampus per 50 μ A of applied skull currents).

Results: Electric fields of as low as 0.35 V/m (0.25–0.47) acutely modulated average firing rate in the hippocampus. At these intensities, firing rate effects increased monotonically with electric field intensity at a rate of 11.5 % per V/m (7.2–18.3). For the majority of excitatory neurons, firing increased for soma-depolarizing stimulation and diminished for soma-hyperpolarizing stimulation. While more diverse, the response of inhibitory neurons followed a similar pattern on average, likely as a result of excitatory drive.

Conclusion: In awake animals, electric fields modulate spiking rate above levels previously observed *in vitro*. Firing rate effects are likely mediated by somatic polarization of pyramidal neurons. We recommend that all future rodent experiments directly measure electric fields to insure rigor and reproducibility.

1. Introduction

The effects of transcranial electric stimulation on neural activity in the brain have been known since the 1960 [1–3]. The acute effects on neuronal firing rate are particularly well established. Namely, the electric fields generated within the brain by transcranial current stimulation can incrementally polarize cell membranes [4] and thus modulate ongoing cell firing [5,6]. The effect acts at the time scale of the neuronal membrane (~30 ms) and thus is relevant for direct current (DC) and most effective for alternating currents (AC) of 30 Hz or less [7, 8]. This acute neuromodulatory effect can be predicted from the orientation and intensity of local electric fields [9,10]. These cellular mechanisms established with *in vitro* animal experiments, also point to network effects [11,12], which can be properly studied only in the intact brain.

However, despite numerous *in-vivo* animal studies in the intervening

decades [12–24], there is still a lack of clarity as to whether the effects observed are clinically relevant, for one simple reason: *in vivo* animal experiments have not adequately characterized electric field magnitudes in the brain. In particular, a significant gap has emerged [25] between electric fields measured *in vivo* in the human brain, which are at or below 0.5 V/m [19,26,27] and field intensities used for *in vitro* animal experiments, which are mostly at or above 5 V/m [28]. Thus, it is difficult to interpret and link results from *in vivo* animal experiments to cellular effects observed *in vitro*. Nor is it clear that the *in vivo* animal experiments have any relevance to the behavioral effects observed in human clinical studies.

To close this gap, we measured electric fields magnitude and their effects on neuronal firing rate *in vivo* in rats and established calibrated computational models of current flow. To do so, we first calibrated our recording equipment on a phantom, and performed *in vivo* field measurements in cortex and hippocampus in a rodent tES model. Then, using

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Conclusion